Modulation of Human Immunodeficiency Virus 1 Replication by Interferon Regulatory Factors

Marco Sgarbanti,1 Alessandra Borsetti,1 Nicola Moscufo,2 Maria C. Bellocci,1 Barbara Ridolfi,1 Filomena Nappi,1 Giulia Marsili,1 Giovanna Marziali,1 Eliana M. Coccia,3 Barbara Ensoli,1 and Angela Battistini1

1Laboratory of Virology, the 2Laboratory of Cellular Biology, and the 3Laboratory of Immunology, Istituto Superiore di Sanità, 00161 Rome, Italy

Abstract

Transcription of the human immunodeficiency virus (HIV)-1 is controlled by the cooperation of virally encoded and host regulatory proteins. The Tat protein is essential for viral replication, however, expression of Tat after virus entry requires HIV-1 promoter activation. A sequence in the 5′ HIV-1 LTR, containing a binding site for transcription factors of the interferon regulatory factors (IRF) family has been suggested to be critical for HIV-1 transcription and replication. Here we show that IRF-1 activates HIV-1 LTR transcription in a dose-dependent fashion and in the absence of Tat. This has biological significance since IRF-1 is produced early upon virus entry, both in cell lines and in primary CD4+ T cells, and before expression of Tat. IRF-1 also cooperates with Tat in amplifying virus gene transcription and replication. This cooperation depends upon a physical interaction that is blocked by overexpression of IRF-8, the natural repressor of IRF-1, and, in turn is released by overexpression of IRF-1. These data suggest a key role of IRF-1 in the early phase of viral replication and/or during viral reactivation from latency, when viral transactivators are absent or present at very low levels, and suggest that the interplay between IRF-1 and IRF-8 may play a key role in virus latency.

Key words: virus replication • Tat • transcription factors • gene expression • T lymphocytes

Introduction

Replication of the HIV-1 provirus is mainly controlled at the transcriptional level and depends on a complex interplay between the viral transregulatory protein Tat and cellular transcription factors with the LTR and the intragenic enhancer in the pol gene (1–4). Several major DNase-hypersensitive sites associated with the LTR promoter and target sequences for regulatory proteins have been identified (5, 6).

In the HIV-1 LTR transcriptional regulatory elements are present both upstream and downstream the transcriptional start site. DNaseI sensitivity studies identified just downstream the 5′ LTR (5) a region spanning at +200 to +217 that is homologous to the IFN-stimulated response element (ISRE)* present in the promoter of IFN-stimulated genes (ISGs; reference 7). This sequence is a binding site for members of the IFN regulatory factor (IRF) protein family (8) and plays a critical role in HIV-1 transcription and replication leading to the definition of a new positive transcriptional regulatory element in the HIV-1 provirus (8, 9).

IRFs play a key role in gene regulation by IFNs and viral infections as well as in several immunological and growth-related cellular functions (10, 11). Nine members of this family have been identified to date based on a homologous DNA-binding domain located at the NH2 terminus responsible for binding to the ISRE. The less conserved COOH-terminal region acts as a regulatory domain and classifies IRFs into three groups: those that activate (IRF-1, IRF-3, IRF-7, and IRF-9/ISGF-3γ), those that repress (IRF-2, IRF-8/ICSBP), and those (IRF-2, IRF-4/LSIRF/Pip) that are able both to activate or to repress gene

*Abbreviations used in this paper: CAT, chloramphenicol acetyltransferase; IRF, IFN regulatory factor; ISG, IFN-stimulated gene; ISRE, IFN-stimulated response element; WB, Western blot.
transcription depending on the target gene. IRFs interact with each other and with other families of transcription factors modifying both ISRE-binding activities and the formation of initiation transcription complexes. In addition IRF-1, IRF-2, and IRF-3 can interact with components of the basal transcriptional machinery as well as with the histone-acetyltransferases (12–14).

The HIV-1 transactivator protein Tat is a ~14/15-kD protein produced early after infection and before virus integration (15), which is absolutely required for productive virus replication (16, 17). Tat has been shown to modulate viral gene expression by increasing the rate of transcription initiation, elongation, and translation of TAR-containing mRNAs (3, 18–20). Several reports also suggest that Tat can dissociate from TAR to bind either elongating RNA polymerase II (21) or DNA-tethered promoter factors (22, 23). Tat has the capability of augmenting transcription of viral as well as cellular genes by both TAR-dependent and TAR-independent mechanisms (24–28) also by acting as a DNA sequence–specific transcription factor in the absence of TAR and other HIV-1 LTR sequences (29).

Both specific and basal cellular transcription factors are key in Tat-mediated transactivation of virus gene expression, including Sp1 (22), TBP (30) and TAFII 55 (31), TAP (32, 33), the kinases TAKs (34), and NF-kB (25). In addition, Tat relieves the transcriptionally inactive chromatin–associated proviral LTR through the recruitment of Tat–associated histone acetyltransferases TAHs (35–37).

The mechanism of action of Tat is complex and not yet completely defined. Similarly, it has not yet been completely elucidated how the viral genome initiates early transcription immediately after viral entry when Tat is still absent or at a threshold concentration, or how the integrated HIV-1 genome reactivates from latency, before viral transactivators are produced.

Here we show that upon entry, HIV-1 is able to induce IRF-1 expression before the expression of Tat. IRF-1 is capable per se de driving LTR-mediated transactivation in a dose-dependent fashion. In addition, IRF-1 increases Tat-mediated transactivation of the HIV-1 LTR via a physical interaction of its COOH-terminal domain with Tat. This positive cooperation is blocked by IRF-8, a physiological repressor of IRF-1 activity, which inhibits Tat-mediated LTR transcription and viral replication in vivo. These results identify IRF-1 as essential for efficient HIV-1 gene expression and viral replication and indicate that the recruitment of IRF-1 to the HIV-1 promoter can be a key step in the early phases of infection or during viral reactivation from latency, in response to both viral infection and cell activation signals.

Materials and Methods

Cell Cultures and Treatments. Jurkat and 293 HEK cells were grown in RPMI 1640 medium (Bio-Whittaker) or MEM, respectively, containing 10% FCS and antibiotics (growth medium). rFDP-γ (Pepro Tech EC LTD) was used at 10 ng/ml. Human PBLs from healthy donors were isolated by Ficoll-Hypaque gradient and the CD4+ T cell population purified by negative selection using magnetic beads (Miltenyi Biotech) coated with mAbs directed against CD8, CD19, CD16, CD56, and CD11b by manufacturers' instructions. The recovered cells were >96% CD3+ as determined by FACS® analysis. Purified cells were cultured in growth medium and activated with anti-CD3 mAb (Clone FM-18; Biosource International).

Plasmids. The HIV-LTR–CAT construct contains the chloramphenicol acetyltransferase (CAT) gene linked to the HIV-1 LTR BH10 clone (~454 to +286) (9). Δ1 LTR corresponds to the BH10-LD1 which is deleted in the ISRE sequence (9). Δ2 LTR and Δ3 LTR were obtained from the HIV-LTR–CAT and Δ1, LTR, respectively, by site-directed mutagenesis of the NF-κB site using the QuickChange site-directed mutagenesis kit (Stratagene). The sequence of the primer used to induce the specific mutation was: 5’ CGAGCTTGCCTACAACCTACCCGGCTGCTCACCAGGGAGG 3’.

CMV-Tat, CMV-IRF-1, CMV-IRF-2, CMV-IRF-3, and CMV-IRF-7* (S477D/S479D) expression vectors have been described previously (38–41); the IRF-8 expression vector (pTarget-ICSBP) was a gift of B. Levi, Technion-Israel Institute of Technology, Haifa, Israel; IRF-4 expression vector was a gift of I. Julkunen, National Public Health Institute, Helsinki, Finland. The pIRF-1/Hygro construct was generated by cloning the fragment excised from pUC-IRF-1 (a gift of T. Taniguchi, University of Tokyo, Tokyo, Japan) by XbaI and HindIII digestion in the pCDNA3.1/Hygro plasmid (Invitrogen Corp.). All plasmids used in the transfection experiments were purified by cesium chloride.

Stable and Transient Transfection Experiments. Stable transfectants of Jurkat cells were obtained by electroporation with a Bio-Rad gene pulser transfection apparatus using a field strength 0.875 kV/cm, a capacitance of 25 μF and a time constant ~10 μs. Cells were selected for 2 wk with 0.5 mg/ml Geneticin G-418 sulfate (GIBCO BRL).

Jurkat cells expressing both IRF-1 and IRF-8 were obtained by transfecting the IRF-8–expressing cells with the pIRF-1/Hygro. After 10 d of selection in growth medium containing 350 μg/ml of Hygromycin (Sigma-Aldrich), cells were amplified on medium containing both hygromycin and geneticin G-418 sulfate.

Bulk populations were frozen and aliquots periodically thawed (every 4–6 wk) to maintain the identity of the polyclonal cell population. Transient transfections experiments were performed using the FuGENE 6 Transfection Reagent (Roche Laboratories) according to the manufacturer’s protocol. Amounts of transfected DNA were normalized by using pCMV vector. A cotransfected RcCMV β-gal plasmid was used to normalize for transfection efficiency.

Enzymatic Assays. CAT assay was performed as described previously (42). β-gal assay was performed using the β-galactosidase Enzyme Assay system (Promega).

EMS4. EMSA with nuclear cell lysates (6 μg; reference 43) was performed as described previously (42). For supershift analysis, nuclear extracts were incubated with polyclonal anti–IRF-1 and anti–IRF-2 (a gift of Dr. J. Hiscott) antibodies in binding buffer containing the oligonucleotide probe (C13 [AACrTGAr4]) for 30 min on ice.

DNA Affinity Purification Assay. A biotinylated oligonucleotide corresponding to the HIV-1RSRE (AGGGACTTGA-AAGCGAAGGAAACCCAGAG) or a mutant oligonucleotide (AGGGACTTGCACCCGGGCAACCGAG) were synthesized (Invitrogen) and then annealed with the corresponding sense oligonucleotides in 1X STE buffer, containing 10 mM Tris-HCl, pH 8.0, 50 mM NaCl and 2 mM EDTA. 25 pico-
moles of biotinylated DNA were mixed with 100–200 µg of nuclear extract in 200 µl of binding buffer containing 20 mM Tris-HCl, pH 7.5, 75 mM KCl, 1 mM DTT, and 5 µg/ml BSA in the presence of 10% glycerol and 20 µg of poly(dI-dC) and incubated for 25 min at room temperature. The complex was pulled down with magnetic beads (Streptavidin MagneSphere; Promega) for 30 min at 4°C and for 10 min at room temperature by mixing with rotation. The collected beads were washed and bound material eluted by boiling in sample buffer. Eluted proteins were separated onto 10% SDS-PAGE followed by immunoblotting with antibody against IRF-1.

**Western Blot Analysis.** Western blot (WB) was performed as described previously (42). Polyclonal antibodies against IRF-1 was a gift of R. Pine, Public Health Institute, New York, NY.

**Immunoprecipitation and Immunoblot Analysis of the Interactions between IRF-1 and Tat.** 293 HEK cells were contransfected with expression plasmids encoding IRF-1 or Tat. Whole cell extracts (200–300 µg) were precleared with rabbit IgG nonimmune antiserum cross-linked to ultralink immobilized protein A-G (Pierce Chemical Co.), and incubated with anti–IRF-1 antibodies (C20; Santa Cruz Biotechnology, Inc.) cross-linked to ultralink immobilized protein A-G sepharose for 1 h at 4°C. Immunoprecipitates were washed five times with lysis buffer and eluted by boiling the beads for 3 min in 1× SDS sample buffer. Eluted proteins were separated by SDS-PAGE and subjected to WB.

**RNA Extraction and Protection Analysis.** Total RNA was isolated by the guanidinium–cesium chloride method (44). RNase protection was performed with 5 µg of total RNA as described previously (42).

To generate the 32P-labeled 280-bp long antisense IRF-1 RNA probe, the pBS-IRF-1 plasmid (45) was linearized with EcoRI and transcribed by T7 polymerase. To generate the 32P-labeled 280-bp long sense IRF-8 RNA probe, the plasmid (pBS-BP) was linearized with PvuI and transcribed by T7 polymerase. The plasmid pBS-BP was generated by cloning a 1,400-bp long fragment obtained by EcoRI digestion from the plasmid pBS-IRF-1 plasmid (45) was linearized with EcoRI and transcribed by T7 polymerase. The plasmid pBS-BP was generated by cloning a 1,400-bp long fragment obtained by EcoRI digestion from the plasmid pTarget-ICSBP containing the entire IRF-8 cDNA. A 18S RNA probe was used as a control for equal RNA loading.

**Virus Stock Preparation and Infection.** Replication-competent T cell-tropic virus was made by transfecting Jurkat cells with the pHXB2K molecular clone, as described previously (46). Viral infection assays were performed by inoculating 106 cells with 1,000 or 5,000 cpm of RT activity corresponding to 0.001 or 0.005 50% TCID50/cell of HIV-1/HXB2. After 2 h, virus was washed out and cells were cultured for 96 h. Virus production was then monitored at 24, 48, 72, and 96 h after infection by measuring the levels of p24 in the culture supernatants with a commercial assay kit (p24/27 core antigen assay; Innogenetics) as specified by the manufacturer.

**RT-PCR Analysis.** To isolate total cellular RNA, 10 × 106 cells were processed using the RNA easy-total RNA extraction kit from QIAGEN. Total RNA was treated with RNase-free DNaseI (Boeringer Mannheim). RT was performed in 50 µl reaction volume containing 1 µg of total RNA according to the manufacturer's instruction (RNA PCR kit; Perkin Elmer). To control for the presence of genomic DNA, control cDNA reaction mixture from which RT was omitted were prepared in parallel. These were uniformly negative (data not shown).

The specific primers named M667, M668, LA45, LA41, M669, and LA23 used to amplify HIV transcripts and PCR conditions were described previously (47). To detect the PCR product of the primer pair M667/M668 and M669/LA23, 32P-labeled primer M669 and M668, respectively, were used for hybridization. Detection of the PCR product using specific primers LA41–LA45 were revealed by hybridization with 32P-labeled oligonucleotide designed to span in between the first and the second Tat exons: TCAAGCAACCCACCTCTCCAA.

To evaluate the expression of the IRF-1 gene, an aliquot of reverse-transcribed-RNA was amplified within the linear range by 25 PCR cycles: denaturation at 94°C for 30 s, annealing at 56°C for 30 s, and extension at 72°C for 30 s. The RT-PCR was normalized for 26S. Each sample was electrophoresed onto 1% agarose, transferred to nylon membrane and hybridized with a specific probe. The following primers and probe for IRF-1 were used: primer 5': GTCCAGCCGAGATGCTAAGAC; primer 3': GCCGTGCCACTCCGACTGCTCC; and probe: GGCCAAAGAGGAAGTCTATGG.

The primers and probe for 26S amplification were: primer 5': GCCCTCAAGATGACAAAG; primer 3': CCAGAAGATAGCCTGTCT; and probe: GAGCGTCTCTCGATGCTATGTTCCCA.

Construction of the Two-Hybrid Clones. The IRF-1 open reading frame (ORF) was PCR-amplified using primers that introduced an EcoRI and an XhoI site at the 5' and 3' ends, respectively, and inserted in the pEG202LexA yeast expression vector in frame with LexA (48). The clones were then transferred in the YEplac181LexA202 vector and used for the yeast two-hybrid interaction assay. The truncated forms IKF-1 (1–291) and IKF-1 (1–234) were constructed by removal of the 3' coding sequences with Accl and BsmI, respectively, followed by the Klenow treatment. For the construction of the VP16-Tat clone, the Tat 86 amino acids open reading frame was PCR-amplified from the CMV–Tat plasmid using primers that introduced an EcoRI and an XhoI site at the 5' and 3' ends, respectively, and cloned in the yeast expression vector pRS314VP16 (49).

The ORFs obtained as described above were cloned into the bacterial expression vector pGEX–4T-1 (Amersham Pharmacia Biotech) at EcoRI and XhoI sites. All the constructs were sequenced before identification.

**Two-Hybrid Yeast Assay.** The two-hybrid yeast assay was performed as described previously (48). Yeast cells harboring a LexA-responsive LacZ reporter plasmid were cotransformed with a LexA/IRF-1 plasmid, and a VP16/Tat plasmid. Transformants were selected at 30°C on YMM plates. From each transformation three colonies were grown at 30°C with 2% galactose, yeast cells were used for the in vitro binding experiments.

In Vivo GST Pull-Down Experiments. GST and GST fusion proteins were expressed in Escherichia coli BL21(DE3)pLysS (48). For the in vitro binding experiments, ~2 µg of GST and GST–Tat or GST–IRF-1 were mixed with the 35S-labeled rIRFs and/or Tat proteins synthesized in vitro using the coupled TNT transcription/translation system (Promega TNT system) in 50 µl of PBS containing 0.1% BSA, and protease inhibitors. Binding reaction was allowed at 4°C for 90 min. Beads were washed, resuspended in sample buffer, and subjected to SDS-PAGE. Gels were analyzed by electronic autoradiography in an Instant Imager (Cambera Packard).

**Results**

IRF-1 Activates Transcription from the HIV-1 LTR and Increases Tat-mediated Transactivation of LTR-directed Gene Expression. The effect of IRFs on HIV-1 transactivation
was evaluated in Jurkat cells transiently cotransfected with vectors expressing IRF-1, IRF-4, or the constitutively activated forms of IRF-3 (IRF-3 5D) and IRF-7 and a HIV-1 LTR-CAT reporter construct (nt -456 to nt +286). As shown in Fig. 1A, the basal activity of the HIV-LTR was increased only by the presence of IRF-1, whereas no or little increase was detected in the presence of the other IRFs.

Therefore, the effect of IRF-1 was further analyzed. IRF-1 increased HIV-1 LTR-directed gene expression in a dose-dependent fashion (Fig. 1B), whereas no activation was detected by deleting the entire COOH-terminal activation domain of IRF-1 (ΔIRF-1). This indicated that upon HIV-1 infection IRF-1 can activate transcription of Tat.

To investigate whether the effect of IRF-1 was mediated by the ISRE, an ISRE-deleted (Δ1 LTR) or a NF-κB mutated (Δ2 LTR) construct were used. As shown in Fig. 1C, IRF-1 was still capable of transactivating the HIV-1 LTR. On the contrary, transactivation was greatly reduced when a mutant bearing deletions in both the ISRE and the NF-κB sites (Δ1 LTR) was used. These results indicate that the ISRE is not the major site mediating the IRF-1 effect.

To determine the effect of the simultaneous presence of IRF-1 and Tat on HIV-1 LTR transactivation, Jurkat cells were cotransfected with the HIV-LTR construct and with both Tat and IRF-1 expression vectors (Fig. 1D). The presence of IRF-1 had additive effects on the HIV-1 LTR-CAT activity induced by suboptimal expression of Tat, whereas the cooperative effect was not evident when Tat was overexpressed (data not shown). This suggests that Tat/IRF-1 effect may be key in the very early phase of infection, when Tat is absent or still at low levels.

HIV-1 induces IRF-1 early upon infection and prior to expression of Tat in both T cell lines and primary CD4+ T cells. To determine whether IRF-1 is induced by HIV-1 and whether this occurs before Tat expression, Jurkat cells were infected with the HIV-1 IIIB strain at a low multiplicity of infection and IRF-1 RNA expression analyzed by RT-PCR at different time points after infection. As shown in Fig. 2A, discrete basal levels of IRF-1 mRNA were detected in Jurkat cells, which increased by 3- and 2.5-fold, respectively, after 5 and 7 h after infection (Fig. 2A and B). This increase was already detectable at 3 h after infection (data not shown) and returned to basal levels within 24 h. A parallel increase in the protein levels was also detected (Fig. 4).

Notably, at the moment of the highest IRF-1 expression (5 h after infection), no doubly/spliced HIV tat/rev RNA transcripts were detected, whereas at 24 h after infection, as expected, the tat/rev mRNA was clearly detectable. Thus, HIV-1 induces IRF-1 expression early upon infection and before production of Tat.

To assess the biological relevance of these findings, experiments were repeated with primary purified CD4+ T lymphocytes stimulated with anti-CD3 mAb and infected with the same virus. IRF-1 and tat/rev RNA expression were then analyzed by RT-PCR at different time points after infection.

As shown in Fig. 3, very low expression of IRF-1 was present in freshly isolated cells, which increased upon stimulation with anti-CD3 antibody (approximately twofold), consistently with previous data (51). However, starting from 5 h after infection IRF-1 mRNA increased by four-

**Figure 1.** Effect of IRFs on HIV-1 LTR transactivation. (A) Jurkat cells were transiently cotransfected with the HIV-1 LTR-CAT (1 μg) and vectors (2 μg) expressing the indicated IRFs. IRF-3 5D and IRF-7* codify for the constitutively activated forms of IRF-3 and -7, respectively (references 40 and 41). After 24 h, CAT activity was evaluated as indicated in the Materials and Methods. (B) Dose–response effect of the wild-type IRF-1 or its mutant deleted in the activation domain (Δ IRF-1) on HIV-1 LTR-directed gene expression. Cells were transfected as in A except that 0.8 μg of HIV-1 LTR-CAT and increasing amounts of the vectors expressing IRF-1 or Δ IRF-1 (0.8, 1.6, 3.2 μg) were used. (C) Effect of IRF-1 on mutated HIV-LTR constructs. Cells were transiently transfected with wild-type HIV-LTR or Δ1, LTR in which the ISRE region is deleted (reference 9), Δ1 LTR in which the NF-κB sites are mutated or Δ2 LTR, in which both sites are deleted/mutated. IRF-1 expressing vector was cotransfected as indicated. Results are shown as percentages of the CAT activity of the wild-type HIV-LTR in IRF-1–transfected cells. (D) IRF-1 cooperates with Tat to enhance HIV-1 LTR-CAT activity. Cells were transiently cotransfected with the HIV-1 LTR-CAT (1 μg), Tat (5 ng), or IRF-1 (1 μg) expressing vectors, either alone or in combination, as indicated. CAT activity was determined after 24 h. The results quantified by an Instant Image are reported as mean levels ± SE from three separate experiments.
fold and its expression peaked at 24 h after infection (sevenfold increase over basal levels). A progressive decrease was then observed as found with Jurkat cells. However, when the IRF-1 stimulation was maximal, no doubly spliced tat/rev RNA transcripts were detected (Fig. 3 C). A similar kinetic of IRF-1 induction was observed with three different healthy donors.

**IRF-1–specific Binding Activity in Infected Cells.** To test whether stimulation of IRF-1 upon HIV-1 infection was associated with the presence of a IRF-1–specific binding activity, EMSA was performed by incubating nuclear cell extracts from infected cells with an oligonucleotide representing the canonical IRF binding site (C13). As shown in Fig. 4 A, discrete complexes were detected both in uninfected and infected cells at all time points after infection, which contained both IRF-1 and IRF-2 or multimers of IRF-2 (52) as anti–IRF-1 and anti–IRF-2 antibodies supershifted the higher-mobility and lower-mobility shifted bands, respectively.

However, early after infection, a specific increase of IRF-1-containing complexes was observed (compare lane 6 versus lane 3), consistent with RNA expression data (Fig. 2). From 7 h after infection onward, IRF-1–specific complexes diminished reaching values comparable or below to those observed in uninfected cells. A representative experiment out of three performed is shown. The increase in IRF-1 mRNA accumulation began to be evident between 5 and 12 h after infection and peaked between 24 and 48 h after infection depending on the donor. 26S RNA was used for normalization as described in Materials and Methods. (B) mRNA relative fold-increase after normalization to the 26S RNA quantified by Instant Imager. (C) Total RNA extracted at the indicated time points as in A was analyzed by RT-PCR for the doubly-spliced (tat/rev) transcript as described in Materials and Methods.
Fig. 4 B, an increasing IRF-1 binding was evident in infected cells at 7 h after infection, which after 24 h returned to basal levels in Jurkat cells but was still present in CD4\(^+\) T cells, according to the RNA expression data (Figs. 2 and 3). This corresponded to the presence of IRF-1 protein (INPUT, Fig. 4 B, right panels) in the same cell extracts. In addition, IRF-1–binding was highly specific since a mutated oligonucleotide (HIV-1-ISRE mut) or an unrelated one (data not shown) did not retain any protein from the same cell extracts.

Taken together, these results demonstrate that IRF-1 and IRF-2 bind to the ISRE-like motif of the HIV-1 LTR. However, early after virus infection, IRF-1 expression increases and this correlates with increasing protein levels and binding to specific LTR-target sequences.

Specific and High-Affinity Binding of IRF-1 and Tat

**GST Pull-Down Experiments.** To determine whether the cooperative effect of Tat and IRF-1 on HIV-1 LTR transactivation (Fig. 1 D) is mediated by physical interactions between the two proteins, GST pull-down assays were performed. IRF-1 was translated in vitro and tested for binding to a GST–Tat fusion protein. As shown in Fig. 5, the GST–Tat protein bound strongly to IRF-1, i.e., up to 30% of the IRF-1 input was bound to the immobilized Tat protein, whereas no binding was detected to control beads containing GST alone. In contrast, IRF-2, IRF-3, IRF-4, IRF-7, and IRF-8 did not bind to Tat. IRF-1 and Tat binding was also detected when a GST–IRF-1 fusion protein was incubated with labeled in vitro–translated Tat protein. In addition, the deletion of the COOH-terminal region of IRF-1 abolished the effect confirming the involvement of this IRF-1 region in binding to Tat. In addition, due to the presence of the transcription activation domain of IRF-1 in the COOH-terminal, dele-
tions of this region also resulted in decreased intrinsic transcriptional activity as compared with the full length IRF-1. Altogether these results indicate that IRF-1 and Tat physically associate and that the COOH-terminal activation domain of IRF-1 is involved in this interaction.

**Binding of Endogenous IRF-1 by Immobilized GST–Tat.** To verify the binding of intracellular IRF-1 with Tat, Jurkat cells were treated or were not treated with IFN-γ for 4 h in order to optimally stimulate IRF-1 expression. Nuclear extracts were then incubated with equal amounts of the GST alone or the GST–Tat fusion protein. After extensive washing, associated proteins were resolved by SDS/PAGE and detected by WB. As shown in Fig. 6 A, a polyclonal antibody specific for IRF-1 detected a major band corresponding to IRF-1 in IFN-γ–induced cells (lane 4) but not in control cells (lane 7), where IRF-1 was only barely detectable. Beads containing a GST–Tat fusion protein were able to selectively bind IRF-1 (lane 6). Conversely, incubation of cell extracts with GST-control beads retained no proteins in controls (lane 8) as well as in cell extracts from IFN-γ–treated cells (lane 5). As control of specificity, the in vitro–translated IRF-1 (lane 1) was incubated with GST–Tat (lane 3).

**Coimmunoprecipitation.** To determine the in vivo interactions between IRF-1 and Tat, coimmunoprecipitation experiments were performed in 293 HEK cells cotransfected with expressing vectors for IRF-1 and Tat. After cotransfection, anti–IRF-1 antibodies were used for immunoprecipitation followed by immunoblot with anti-Tat antibodies. As shown in Fig. 6 B, the Tat protein was readily detected in the anti–IRF-1 immunocomplexes obtained from cells cotransfected with both RcCMV/IRF-1 or RcCMV/Tat, whereas Tat was not detected in cells transfected with RcCMV/IRF-1 or RcCMV/Tat alone (Fig. 6 B). Vice versa, when immunoprecipitation was performed with anti–Tat antibodies followed by immunoblot with anti–IRF-1 antibodies, the IRF-1 protein coimmunoprecipitated with Tat in cells extracts of doubly transfected cells (data not shown). Altogether these results indicate that IRF-1 and Tat associate intracellularly.

**Table I.** In Vivo Interaction between IRF-1 and Tat by the Two-Hybrid System

<table>
<thead>
<tr>
<th>VP16</th>
<th>VP16–Tat</th>
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<tr>
<td>LexA</td>
<td>0.54 ± 0.1 0.53 ± 0.03</td>
</tr>
<tr>
<td>LexA–IRF-1</td>
<td>581 ± 27  819 ± 117</td>
</tr>
<tr>
<td>LexA–IRF-1 (1–291)</td>
<td>453 ± 80  539 ± 105</td>
</tr>
<tr>
<td>LexA–IRF-1 (1–234)</td>
<td>131 ± 65  121 ± 10</td>
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Yeast strain DBY1 was transformed with the yeast expression vectors Yeplac181G (Lex2) containing LexA or LexA–IRFs sequences, pH5314 (Trp1) containing VP16TAD or VP16TAD–Tat, and the LacZ reporter vector pSH18 (Ura3). Transformants were selected and analyzed as described in Materials and Methods. β-galactosidase activity is expressed as mean β-galactosidase units ± SD.

**Figure 6.** IRF-1 and Tat associate intracellularly. (A) In vitro–translated IRF-1 (lanes 1–3) and nuclear cell extracts from Jurkat cells treated with IFN-γ (lanes 4–6) or control medium (lanes 7–9) were incubated with purified GST–Tat fusion protein or GST alone. Bound proteins were then analyzed by WB using anti–IRF-1 polyclonal antibody as described in Materials and Methods. 10% of the extract used for binding assays is shown in lane 7 (untreated cells) and lane 4 (IFN-γ–treated cells). The slowly migrating IRF-2 protein band observed in IFN-γ–treated cell extracts is due to the phosphorylation induced by IFN-γ (reference 64). Extra bands in lanes 7–9 are not specific. (B) 293 HEK cells were transfected with the expression plasmids encoding IRF-1 or Tat, alone or in combination. Whole cell extracts (300 µg) were immunoprecipitated with anti–IRF-1 antibodies (αIRF-1). Immunoprecipitated complexes were separated by 10% SDS-PAGE and subsequently probed with anti–Tat antibodies (αTat) as indicated. Whole cell extracts (10 µg) were separated on 10% or 15% SDS-PAGE and probed with anti-Tat or anti–IRF-1 antibodies.
the IRF-8 inhibitory effect, Jurkat cells were stably transduced with IRF-8 and then transfected with the HIV-LTR CAT construct and the Tat-expression vector. After selection, transgene expression was assessed by RNase protection assay (Fig. 7 C). Bulk populations of transfected cells were chosen to avoid clonal variability. As shown in Fig. 7 D, the constitutive expression of IRF-8 reduced by ∼50% both the IRF-1 and the Tat-directed HIV-1–LTR transcription.

**Inhibition of HIV-1 Replication in IRF-8–expressing Cells.** To evaluate the inhibitory effect of IRF-8 on virus replication, Jurkat cells stably expressing IRF-8 or control cells containing the vector alone were infected with 1,000 and 5,000 cpm/ml, corresponding to 0.001/0.005 TCDI50 per cell of the HIV-1 IIIB strain virus. The accumulation of HIV-1 RNA species was then evaluated by semiquantitative RT-PCR (Fig. 8 A) at 24 and 48 h after infection. In control cells (lanes 1, 3, and 5), all HIV transcripts (unspliced, singly- or multi-spliced) were clearly detected after 2 d of infection. In contrast, in IRF-8–expressing cells (lanes 2, 4, and 6), a significant decrease of both spliced and unspliced viral RNA was observed at both infection doses, the doubly-spliced tat/rev being the more reduced. This correlated with an abolished or a reduced virus replication (Fig. 8 B). Specifically, at a low multiplicity of infection, p24 antigen production was, only barely detectable at 48 h after infection and undetectable at later time points, as compared with control cells. Similarly, a reduction of >3 logs was progressively observed, in cells infected with 5,000 cpm/ml of virus. This indicates that IRF-8 represses HIV-1 productive infection.

**IRF-8 Blocks Activation of HIV-1 LTR Transcription by Interfering with IRF-1–Tat Binding.** Since no direct activity of IRF-8 on LTR transcription nor a direct binding of IRF-8 to Tat was detected (Figs. 5 and 7), to investigate the molecular mechanism(s) responsible for IRF-8–mediated inhibition of HIV, GST–pull down assays were performed with in vitro–translated IRF-1 and IRF-8, and the labeled proteins were tested for binding to immobilized Tat (Fig. 9). The binding of IRF-1 to immobilized Tat was clearly impaired in the presence of IRF-8, since the IRF-1 input retained on the GST–Tat beads was reduced by ∼50% (compare lanes 2 and 3). On the other hand, the presence of IRF-2 did not affect the IRF-1 binding to immobilized Tat (compare lanes 3 and 4). These results indicate that the inhibitory effect exerted by IRF-8 is, at least in part, mediated by the competition of IRF-8 and Tat for the binding to IRF-1.

To further support this conclusion, Jurkat cells expressing IRF-8 were stably transduced also with an IRF-1–expressing vector. After selection, transgene expression was assessed by RNase protection analysis with RcCMV/IRF-8 cells (control) and with RcCMV/IRF-8/IRF-1 doubly transfected cells (Fig. 10 A). A polyclonal population of cells overexpressing both IRF-8 and IRF-1 was then infected with 1,000 and 5,000 cpm/ml and the production of the HIV p24 antigen evaluated. The dramatic reduction of p24 accumulation in IRF-8 constitutively expressing cells...
was reversed by at least 50–70% in the cells overexpressing also IRF-1 (Fig. 10 B). Thus, IRF-1 overcomes the inhibitory effect of IRF-8 on HIV-1 replication.

Discussion

HIV-1 transcription and replication is controlled by both viral and cellular factors, which act at the transcriptional, posttranscriptional, and/or translational levels. Both basal- and tissue-specific transcription factors that are essential for HIV-1 have been identified. In the present report we examined the role of the cellular IRFs in HIV-1 transcription and replication and their interactions with the viral transactivator Tat.

We identified IRF-1 as an essential factor for efficient HIV-1 gene expression especially in the early phase of viral replication and before expression of Tat. Several lines of evidence support this conclusion: (i) IRF-1 activates LTR-driven transcription in the absence of the viral transactivator Tat; (ii) IRF-1 is induced at very early time after virus infection and before expression of Tat; (iii) IRF-1 expression during infection correlates with a specific binding to the ISRE of the HIV-1 LTR; (iv) in the presence of low doses of Tat, IRF-1 increases Tat-mediated HIV-1 transactivation by a direct physical interaction with Tat through its transactivation domain; (v) IRF-8, a dominant negative regulator of IRF-1 activity, blocks HIV-1 transcription both in vitro and in vivo, and inhibition is released by overexpression of IRF-1.

The role of IRFs in the regulation of IFN and ISGs (54–56), as well as of genes expressed during inflammation, immune responses, hematopoiesis, cell proliferation, and differentiation has been clearly defined (10, 11). The recent identification of an ISRE on the HIV-1 LTR downstream the transcription start site together with the demonstration that sequences comprising the ISRE are essential for efficient HIV-1 transcription and virus replication (8, 9), allowed us to speculate that IRFs exert a role in HIV-1 transcription. Indeed we demonstrated that IRF-1, but not other IRFs, activates the HIV-1 promoter. IRF-1 may, thus, effectively activate transcription of Tat and, in turn, amplify HIV-1 transcription and virus replication. This can be particularly relevant at the initial phases of HIV-1 replication when viral transactivators are not yet synthesized or are present at subthreshold concentrations.

This has biological significance since IRF-1 is stimulated early after virus infection and before expression of Tat in both cell lines and primary CD4+ T lymphocytes. The kinetic of IRF-1 induction closely resembles that described in cells infected by the vesicular stomatitis virus or Newcastle disease virus, where IRF-1 expression precedes IFN
and virus replication. Therefore, we propose that IRF-1 exerts a key role in initiating and amplifying transcription from the HIV-1 LTR, increasing production of Tat, which, in turn, thereby amplifies LTR-directed gene expression. Consistent with this, IRF-1 binds to Tat and cooperates with suboptimal doses of Tat to activate transcription (Figs. 1 and 5). Of note both Tat and IRF-1 have been shown to functionally interact with general transcription factors such as TFIIB (12, 33) and coactivators or adaptors, such as the histone acetyltransferases p300/CBP and pCAF (13, 35–37). These interactions occur through different domains of the proteins thus, IRF-1 might modulate the HIV-1 LTR promoter activity also by acting as a bridge between Tat and component(s) of the basal transcriptional machinery and/or may participate to the Tat-holoenzyme complex according to the model proposed by Cujec et al. (23). It remains to clarify at what extent IRF-1 is critical for HIV-1 replication in T cells. To this purpose a strategy leading to inhibition of IRF-1 expression both basal- and virus-induced, but not interfering with cell viability, should be investigated.

In addition to activate HIV gene expression and replication the IRF system can also repress it specifically, since we showed that IRF-8 is able to impair the binding of IRF-1 to Tat in vitro and to drastically reduce HIV-1 replication in vivo and that this block is released by the simultaneous overexpression of IRF-1. It is, thus, conceivable that, since IRF-8 does not contain an activation/repression domain, excess of IRF-8 compressing IRF-1 may inhibit the IRF-1–induced LTR transcription and the binding of IRF-1 to Tat further impairs HIV-1 replication ultimately leading to a block of viral replication. These data, therefore, suggest that an increase in IRF-8 expression can be involved in the establishment of latency, whereas activation of IRF-1 expression functions as a positive regulator of HIV-1 transcription and replication. Thus the differential expression of these IRFs in activated versus non activated cells and in different cell types may determine productive infection and/or virus reactivation at different tissue sites.

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